





## Dorema Aucheri and exercise training in enhancing PPAR $\gamma$ /mTOR/PI3K gene expression in muscle tissue of hypothyroid mice: Experimental approaches using a hypothyroid mouse model

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### Abstract

**Background:** Hypothyroidism is typically associated with a decreased basal metabolic rate, reduced energy expenditure, and weight gain. Exercise training and Dorema Aucheri (DA) have been identified as beneficial therapeutic strategies within complementary health approaches. Skeletal muscle metabolism and fiber type are regulated by innervation and soluble factors, such as thyroid hormones. However, the mechanisms between muscle function and hypothyroidism remain unclear.

**Methods:** Thirty mice were divided into five subgroups: the normal group; hypothyroid mice (HYPO, 8 mg/kg of propylthiouracil administered via intraperitoneal injection for 30 days); hypothyroid mice treated with DA (Gavaged at 0.4 mg/kg for two months, five days per week); hypothyroid mice treated with exercise (75% VO<sub>2</sub> max, 45 minutes per session, for two months, five days per week); and hypothyroid mice treated with both DA and exercise. The mRNA expression levels were detected via real-time qPCR.

**Results:** The data indicated that PPAR $\gamma$ , mTOR, and PI3K levels are reduced in hypothyroidism. DA and exercise enhanced PPAR $\gamma$ , mTOR, and PI3K levels in muscle tissue. Notably, DA and exercise significantly increased the expression levels of PPAR $\gamma$ , mTOR, and PI3K.

**Conclusion:** Exercise and DA, as alternative and complementary medicine, modified the PPAR $\gamma$ /mTOR/PI3K signaling pathways affected by hypothyroidism in mice.

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### Highlights

#### What is current knowledge?

- Exercise training may ameliorate the PPAR $\gamma$ /mTOR/PI3K signaling pathway
- DA can regulate PPAR $\gamma$ /mTOR/PI3K expression in hypothyroidism.

#### What is new here?

Exercise training and DA regulated the metabolism and atrophy signaling pathways in the muscle tissue of hypothyroid mice.

### Introduction

Thyroid hormones (THs), specifically tri-iodothyronine (T<sub>3</sub>) and thyroxine (T<sub>4</sub>), play a vital role in the growth and maturation of the central nervous system, lungs, and skeletal muscle (1,2). Thyroid-stimulating hormone (TSH) promotes the synthesis and release of THs; however, THs adversely regulate TSH through a feedback loop (2). Skeletal muscle metabolism and fiber type are controlled by innervation and soluble factors, such as THs (3). Hypothyroidism is a condition caused by an underactive thyroid gland, resulting in insufficient production of THs essential for regulating metabolism and overall bodily functions. Common symptoms include fatigue, weight gain, cold intolerance, and depression, which can sometimes be subtle and lead to underdiagnosis (4). This condition is particularly prevalent in women and older adults; in the U.S., approximately 4.6% of the adult population is affected, with higher rates observed in women over 60. If left untreated, hypothyroidism can lead to serious complications such as heart disease, infertility, and severe depression, highlighting the significance of awareness, early diagnosis, and effective management strategies (5,6).

Muscle growth, homeostasis, and regeneration require the binding of T<sub>3</sub> to thyroid hormone nuclear receptors (7). Studies have shown that the T<sub>3</sub> hormone affects skeletal muscle metabolism and neutralizes muscle atrophy (8). A sufficient serum level of THs is essential for skeletal muscle homeostasis, as muscle function is impaired in both hypothyroid and hyperthyroid conditions (9,10). Skeletal muscle is a crucial component of the body, influencing quality of life, health, survival, and metabolic balance (11). It serves as the body's primary protein reservoir and plays a vital role in regulating glucose and lipid homeostasis (12). Consequently, changes in muscle mass, whether growth or loss, can affect general metabolism, movement, eating, and breathing (13). Research has demonstrated that thyroid dysfunction causes muscle atrophy (14-16).

T<sub>3</sub> significantly influences skeletal muscle metabolism and helps counteract muscle atrophy through various mechanisms. By binding to nuclear thyroid hormone receptors, T<sub>3</sub> regulates gene expression, promoting the production of

anabolic proteins while inhibiting catabolic ones (17). It enhances protein synthesis by activating the mTOR pathway, facilitates the conversion of muscle fibers to more metabolically active fast-twitch types, and increases the uptake of glucose and fatty acids for energy (18). Additionally, T<sub>3</sub> may reduce myostatin levels, a protein that inhibits muscle growth. Together, these actions contribute to the preservation and growth of muscle, underscoring T<sub>3</sub>'s importance in maintaining skeletal muscle health (12). In conclusion, hypothyroidism is a complex disease characterized by various physiological symptoms that can significantly impair quality of life, including a decrease in physical performance (19).

The essential oil derived from the above-ground sections of Dorema Aucheri (DA) contains a high concentration of flavonoids and terpenoids (20). However, growing evidence indicates that phytochemicals impact THs and antioxidant enzymes (21). Phytochemical products are valuable substances with diverse applications in daily life (22). These natural compounds may serve as complementary and alternative medicines to help prevent and alleviate disease symptoms, aligning with traditional medical practices (23). According to a recent study, the essential oil derived from the aerial parts of DA is rich in flavonoids and terpenoids, which grant it antioxidant, anti-tumor, anti-diabetic, anti-hypercholesterolemic, anti-hyperlipidemic, and anti-inflammatory properties (24).

Additionally, evidence suggests that combining phytochemicals influences THs, sex hormones, and antioxidant enzyme activity. Given the importance of safety and efficacy in herbal medicine, this study evaluates the effectiveness of the phytochemical cocktail through a computational phytochemistry analysis focused on potential cutpoints in disease pathogenesis (25). Mostafavi et al. demonstrated that DA at a dosage of 0.4 mg/kg protected liver function in a mouse model, compared to higher doses of 0.8 mL/kg, 1.6 mL/kg, and 3.2 mL/kg (24).

Physical activity and exercise training have been defined as beneficial therapeutic strategies in complementary health approaches (26,27). Prolonged exercise training can significantly impact bodily performance, metabolism, and hormonal activity (28). Another key player in regulating metabolism is the peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ )/peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1 $\alpha$ ) axis. Evidence has demonstrated that exercise training modifies thyroid hormone levels and changes the relative expression of PPAR $\gamma$  (29). PPAR $\gamma$  expression is induced by regular exercise training (30). Huh et al. have shown that plasma PPAR $\gamma$  gene expression increases in response to exercise (31). Importantly, there is a positive association between mTOR and PI3K in trained mice (32).

The aim of this study is to evaluate the therapeutic effects of DA and exercise training on thyroid hormone levels and the expression of the PPAR $\gamma$ /mTOR/PI3K signaling pathways in skeletal muscle hypothyroid mice. It seeks to understand

how DA and exercise training may influence the physiological processes regulated by THs and the expression levels of the PPAR $\gamma$ /mTOR/PI3K signaling pathways in skeletal muscle hypothyroid mice.

## Methods

### Ethical code

This study was conducted and authorized following the laboratory animal protocols approved by the Islamic Azad University Ethics Committee, Khorasgan Branch (IR.IAU.KHUISF.REC.1400.231).

Thirty six-week-old C57BL/6 mice with an approximate weight of  $19 \pm 2$  g were purchased from the Royan Institute. They were maintained in the animal house of the Isfahan (Khorasgan) Branch of Islamic Azad University under controlled environmental conditions (Temperature of  $23 \pm 4^\circ\text{C}$ ,  $60\% \pm 4\%$  humidity, and a 12-hour light/dark cycle). After one week of adaptation, the mice were separated into two groups: the control group (Mice with no treatment) and the hypothyroidism group, which was injected daily with 8 mg/kg of propylthiouracil (PTU) for one month. The sample size of thirty mice (Six mice per group) was estimated using G\*POWER software and a literature review.

It should be noted that in this study, the sample replication was considered as biological replicates. Before assigning the mice to groups, they were typically numbered or tagged for identification. Mice were randomly assigned to groups using a computer program or random number tables. Each mouse was assigned to a subgroup based on the chosen randomization method to ensure that every group had a similar baseline of variables, thus enhancing the reliability of the study results. Therefore, the mice were divided into five subgroups (Each group,  $n = 6$ ): 1. A control group, 2. Mice with hypothyroidism (HYPO), 3. Mice with hypothyroidism treated with DA at a dosage of 0.4 mg/kg (HYPO-DA), 4. Mice with hypothyroidism treated with aerobic exercise (HYPO-AE), and 5. Mice with hypothyroidism treated with both DA and aerobic exercise (HYPO-DA+AE). The research employed minimally invasive techniques for administering treatments and collecting data to reduce trauma and discomfort to the animals. Moreover, anesthetics were used where appropriate to manage pain and distress during procedures, ensuring the animals experienced as little suffering as possible. Furthermore, continuous monitoring of the animals was conducted throughout the study to assess their well-being, allowing for the timely identification and alleviation of any signs of pain or distress.

### Experimental induction of hypothyroidism in mice

Based on the literature review, we selected 8 mg/kg of PTU as the optimal dose to induce hypothyroidism in mice (33). In this study, PTU was administered intraperitoneally at 8 mg/kg for 30 days. During the treatment with PTU, several nonspecific clinical signs were noted in the affected mice. Specifically, while inducing hypothyroidism with PTU, we observed reduced skin turgor, a roughened coat, a hunched posture, and/or decreased activity. Notably, this study did not record any morbidity related to the experimental procedures.

### Consuming a mixture of Dorema Aucherii

DA was dried in a darkroom and then powdered using an electric mixer, following the hydrodistillation methodology described in earlier investigations to extract bioactive components. In this study, we administered DA at a 0.4 mg/kg dosage for two months and five days (34).

### Aerobic exercise protocol

For eight weeks (5 days/week), mice with hypothyroidism underwent aerobic training on a motorized treadmill. The aerobic exercise performed was moderate to high-intensity interval training. Following one week of adaptation, the training protocol's duration, intensity, and repetitions were progressively increased until reaching approximately 75% of the maximum oxygen consumption (VO<sub>2</sub> max) at 45 minutes and a 0 slope. The workout regimen was designed to increase in intensity and speed gradually. The rate of increase was set at 3 m/min and 5% of VO<sub>2</sub> max. The protocol started at 10 m/min and 20% VO<sub>2</sub> max and progressed to 25 m/min and approximately 75% VO<sub>2</sub> max (35). The aerobic exercise protocol is indicated in Table 1.

Table 1. The aerobic exercise protocol

| Week       | Speed (m/min)                  | Exercise Duration/Session (min) | Exercise Sessions/Week |
|------------|--------------------------------|---------------------------------|------------------------|
| Adaptation | 7-10                           | 10                              | 5                      |
| First      | 7, 10, 13, 10, 7               | 10, 3, 19, 3, 10                | 5                      |
| Second     | 7, 10, 13, 15, 13, 10, 7       | 5, 3, 3, 23, 3, 3, 5            | 5                      |
| Third      | 10, 13, 16, 17, 16, 13, 10     | 5, 3, 3, 23, 3, 3, 5            | 5                      |
| Fourth     | 10, 13, 16, 18, 16, 13, 10     | 5, 3, 3, 23, 3, 3, 5            | 5                      |
| Fifth      | 10, 13, 16, 19, 16, 13, 10     | 5, 3, 3, 23, 3, 3, 5            | 5                      |
| Sixth      | 10, 13, 16, 20, 16, 13, 10     | 5, 3, 3, 23, 3, 3, 5            | 5                      |
| Seventh    | 10, 13, 16, 18, 21, 16, 13, 10 | 5, 3, 3, 3, 20, 3, 3, 5         | 5                      |
| Eighth     | 10, 13, 16, 19, 22, 16, 13, 10 | 5, 3, 3, 3, 20, 3, 3, 5         | 5                      |

### Measurement of hormone levels

Blood collection was performed under anesthesia to minimize distress. The operator inserted a needle directly into the heart through the chest wall to collect blood. The serum was then separated using a centrifuge at a speed of 3100 g for

5 minutes at a temperature of  $4^\circ\text{C}$ . The concentration levels of T3 (MYBIOSOURCE, MBS262762) were measured to confirm the presence of hypothyroidism.

### RNA extraction and qPCR-Real Time

The muscle tissue samples were subjected to RNA extraction using Trizol reagent (Thermo Scientific, USA) according to the manufacturer's instructions. The RNA quality was assessed using NanoDrop spectrophotometry (Thermo Scientific, Waltham, MA, USA). Next, cDNA synthesis was performed according to the manufacturer's protocol provided by TaKaRa. mRNA primers were designed using Oligo 7 and Beacon Designer 7 software. In addition, real-time qPCR was performed using SYBR Green dye (TaKaRa, Kusatsu, Japan) on a Rotor-Gene 6000 apparatus (Corbett Life Science, Mortlake, Australia). The study utilized 18S ribosomal RNA (18S rRNA) as a reference gene to standardize the expression of genes (Table 2).

Table 2. The sequences of primers used for real-time PCR

| Genes            | Sequences (5'- 3')     |
|------------------|------------------------|
| PPAR $\gamma$ -f | CTGTTATGGGTGAAACTCTGG  |
| PPAR $\gamma$ -r | GTGGTAAAGGGCTTGATGTC   |
| PI3K-f           | ACACCACGGTTTGGACTATGG  |
| PI3K-r           | GGCTACAGTAGTGGGCTTGG   |
| mTOR-f           | AGAAGGGTCTCCAAGGACGACT |
| mTOR-r           | CTCCAAGGCACTTGACTGCTG  |
| 18S rRNA-f       | CGGACACGGACAGGATTG     |
| 18S rRNA-r       | TCGCTCCACCAACTAAGAAC   |

### Statistical analysis

The analysis of variances was computed using GraphPad Prism Software (Version 9, GraphPad Software Inc., La Jolla, CA). In addition, the Shapiro-Wilk test was employed to assess the normality of the distribution in this study. Furthermore, all data were analyzed using one-way analysis of variance (ANOVA) with Tukey's post hoc test and t-test. The results are presented as the mean value  $\pm$  standard deviation (SD). Differences with a P-Value  $< 0.05$  are considered statistically significant in all analyses.

## Results

### The T3 hormone changed by exercise training and Dorema Aucherii

The concentration of T3 hormone in hypothyroid mice was significantly different compared to the control group (Table 3, P-Value  $< 0.001$ ). Based on the data, T3 concentration was reduced in the HYPO group compared to the Control group (Table 3, P-Value  $< 0.001$ ). In addition, it was observed that the concentration of T3 in the HYPO-DA and HYPO-AE groups was higher compared to the HYPO group (Table 3, P-Value  $< 0.001$ ). Furthermore, the concentration of T3 significantly increased in the HYPO-DA+AE group compared to the other groups (Table 3, P-Value  $< 0.001$ ).

Table 3. The T3 concentration

| Groups     | T3 concentration (ng/ml)        | P-Value     |
|------------|---------------------------------|-------------|
| Control    | 7.31 $\pm$ 0.64                 | 0.001 $< P$ |
| HYPO       | 1.2 $\pm$ 0.16 <sup>a</sup>     | 0.001 $< P$ |
| HYPO-DA    | 3.64 $\pm$ 0.65 <sup>ab</sup>   | 0.001 $< P$ |
| HYPO-AE    | 3.12 $\pm$ 0.39 <sup>ab</sup>   | 0.001 $< P$ |
| HYPO-DA+AE | 5.64 $\pm$ 0.98 <sup>abcd</sup> | 0.001 $< P$ |

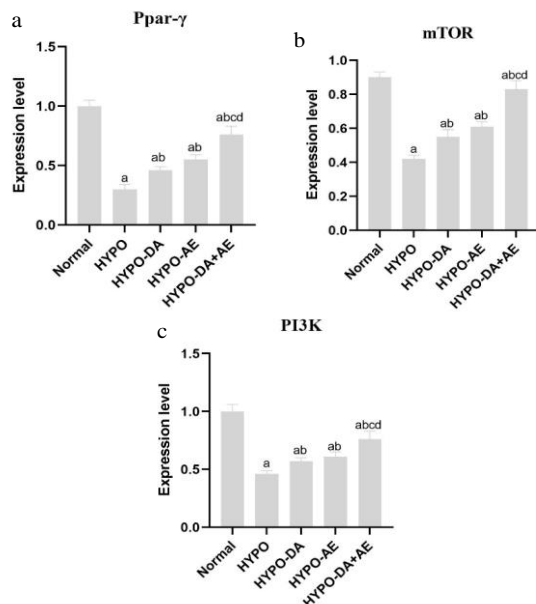
All data are presented as mean  $\pm$  SD. "a" indicates the significance between the normal and other groups. "b" indicates the significance between the HYPO and other groups. "c" indicates the significance between HYPO-DA and HYPO-DA+AE. "d" indicates the significance between HYPO-AE and HYPO-DA+AE.

### The expression level of the PPAR $\gamma$ /mTOR/PI3K enhanced by exercise training and Dorema Aucherii

The PPAR $\gamma$ /mTOR/PI3K expression level in the HYPO group significantly decreased compared with the control group (Figure 1a-c, P-Value  $< 0.001$ ). Our data showed that DA increased PPAR $\gamma$  gene expression in the muscle by 0.16-fold (Figure 1a, P-Value  $< 0.001$ ). Moreover, exercise training enhanced PPAR $\gamma$  gene expression in the muscle by 0.25-fold (Figure 1a, P-Value  $< 0.001$ ). It should be noted that the relative expression of PPAR $\gamma$  was amplified in the muscle of the HYPO-DA+AE group by 0.46-fold (Figure 1a, P-Value  $< 0.001$ ).

PTU reduced the relative expression of mTOR in the muscle of HYPO mice by 0.42-fold (Figure 1a, P-Value  $< 0.001$ ). The mTOR expression level was significantly increased by DA (0.1-fold) and aerobic exercise (0.19-fold) (Figure 1b, P-Value  $< 0.001$ ). In addition, the mTOR gene was elevated in the muscle of the HYPO-DA+AE group by 0.41-fold (Figure 1b, P-Value  $< 0.001$ ).

The PI3K expression level in the muscle of HYPO mice was reduced by 0.46-fold (Figure 1c, P-Value  $< 0.001$ ). The PI3K gene was significantly amplified by DA (0.11-fold) and aerobic exercise (0.16-fold) (Figure 1c, P-Value  $< 0.001$ ). Furthermore, the PI3K gene was raised in the muscle of the HYPO-DA+AE group by 0.3-fold (Figure 1c, P-Value  $< 0.001$ ). Notably, we found that the relative expression of PPAR $\gamma$ /mTOR/PI3K was improved in the HYPO-DA+AE group (Figure 1a-c, P-Value  $< 0.001$ ).



**Figure 1.** The Dorema Aucheri and exercise training regulated PPAR $\gamma$ /mTOR/PI3K pathway. a. the relative expression level of PPAR $\gamma$ . b. the relative expression level of mTOR. c. the relative expression level of PI3K. All data are shown as mean  $\pm$  SD. a indicates the significance between the normal and other groups. b indicates the significance between the HYPO and other groups. c indicates the significance between HYPO-DA and HYPO-DA+AE. d indicates the significance between HYPO-AE and HYPO-DA+AE.

## Discussion

The present study showed that hypothyroidism in mice impaired the PPAR $\gamma$ /mTOR/PI3K pathway. Our results showed that the PPAR $\gamma$ /mTOR/PI3K expression level significantly decreased in hypothyroidism in mice. Moreover, we demonstrated that the PPAR $\gamma$ /mTOR/PI3K pathway was dysregulated. Furthermore, we revealed that the expression level of PPAR $\gamma$ /mTOR/PI3K was reversed by exercise training and DA.

Skeletal muscles are one of the important components of the body that are related to quality of life, health, survival, and metabolic balance (11). Skeletal muscle is the protein reservoir of our body and an essential regulator of glucose and lipid homeostasis (11,36). As a result, the growth or loss of muscle mass can affect general metabolism, movement, eating, and breathing (13). Therefore, it is not surprising that excessive muscle loss is a poor prognostic indicator for a variety of diseases, including cancer, organ failure, infections, and unhealthy aging (37). Muscle function is influenced by different qualitative systems that regulate the function of proteins and contractile organelles (38). It is worth mentioning that the expression of genes in skeletal muscle can depend on THs. MCT8 and MCT10, thyroid hormone transporters, and thyroid hormone converting enzymes, DIO2 and DIO3, are expressed in the skeletal muscle of humans and rodents (39).

We found that consuming DA could improve the PPAR $\gamma$ /mTOR/PI3K expression level in hypothyroid mice. DA contains calcium and magnesium, which can produce and increase TSH as a mediator of the second messenger through the calcium-phosphatidyl-inositol mechanism (40). In addition, coumarin is one of the compounds in DA that affects thyroid function by inhibiting the conversion of T4 to T3 (41). It also contains many antioxidant agents, such as flavonoids, anthocyanins, and phenolic acid (42).

Our data indicated that the expression level of mTOR changed in the hyperthyroid condition. Coppola et al. established that in a rat model with central hyperthyroidism, the activation of the mammalian target of rapamycin (mTOR) signaling in the arcuate nucleus (ARC) of the hypothalamus enhanced feeding (43). Additionally, Coppola et al. demonstrated that fasting stimulates the local production of T3 in the ARC, which promotes feeding by increasing UCP2-mediated mitochondrial uncoupling (43). It can be deduced that the converse happens in hypothyroidism, as Coppola et al. have documented reduced levels of hypothalamic Ucp2 mRNA under hypothyroid conditions. Moreover, there is a correlation between heightened SNS signaling and lower food intake (44,45). Evidence has demonstrated that mTOR signals in the central circadian clock, located in the suprachiasmatic nucleus (SCN) of the hypothalamus, control the production of clock proteins Period 1 and Period 2, thereby modifying circadian rhythms. As previously stated, THs control mTOR signaling in the ARC to enhance food intake; thus, it is reasonable to assume that hypothyroidism affects mTOR signaling in the suprachiasmatic nucleus (SCN) (46). Nevertheless, a comprehensive investigation is still required to determine the mechanisms by which TH diseases induce metabolic and behavioral alterations by modifying circadian rhythms (47).

DA, a traditional herbal plant, has gained attention for its potential effects on thyroid function (24). It has been noted for its ability to modulate thyroid activity, which may be beneficial in both hyperthyroidism and hypothyroidism by supporting the regulation of THs (24). Calcium ions are crucial in numerous cellular processes, including hormone secretion. Phosphatidylinositol (PI) is a phospholipid in cell membranes that can be phosphorylated to generate inositol trisphosphate (IP3). This process is essential for mobilizing intracellular calcium. Increased intracellular calcium levels can promote the secretion of THs by stimulating the exocytosis of thyroid follicle cells. Enhanced calcium signaling can increase the synthesis and release of THs (T3 and T4), improving metabolic functions and energy regulation. Coumarins are a class of compounds found in various plants, including DA (48). They possess anti-inflammatory, antioxidant, and potential endocrine-modulating properties. Some studies suggest that coumarin compounds may influence thyroid function by interacting with thyroid hormone receptors or pathways involved in their synthesis. Improvements in PPAR $\gamma$ /mTOR/PI3K expression could signify that DA facilitates favorable environments for thyroid hormone synthesis and secretion through enhanced energy metabolism and cellular signaling pathways. The modulation of calcium levels via phosphatidylinositol and the influence of coumarins may support these pathways, improving thyroid health and function.

In this study, we found that the relative expression of PPAR $\gamma$ /PI3K was improved by exercise training. Exercise can regulate the amount of hormones secreted by the thyroid gland. Furthermore, since exercise regulates metabolism in muscle tissue over the long term, it can have a dual effect on the function of the thyroid gland and the locomotor system. By increasing THs, exercise regulates both bone metabolism and muscle function, thereby improving hypothyroidism. Implementing specific exercise protocols can affect the regulation of thyroid hormone levels over the long term and positively improve the function of other body tissues, including bone and muscle (49). It is well-known that exercise affects the activity of many glands and their hormone production (49), and increasing blood circulation leads to improved thyroid hormone production and body metabolism (50). On the other hand, exercise also leads to the secretion of epinephrine, cortisol, growth hormone, prolactin, and other factors that modulate the immune system (51).

## Conclusion

It seems that the use of medicinal plants, along with exercise training, can have a better protective effect on hypothyroidism in mice. According to the results of the present study, DA and aerobic exercise modified the PPAR $\gamma$ /mTOR/PI3K signaling pathways induced by hypothyroidism in mice.

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## Ethical statement

All animal studies were performed following the standard ethical guidelines for working with laboratory animals, approved by the Azad University of Isfahan (Khorasgan, Iran), and after receiving the code of ethics (IR.IAU.KHUISF.REC.1400.231).

## Conflicts of interest

The authors declare that they have no conflict of interest.

## Author contributions

MB: carried out the experiments and drafted the manuscript. FT: participated in this study as the supervisor and contributed to the study design, conceptualization, data analysis, validation, and manuscript revision. KHJD and RM: participated in data validation and manuscript revision. All authors read and approved the final manuscript.

## References

- Silva JE. Thermogenic mechanisms and their hormonal regulation. *Physiol Rev.* 2006;86(2):435-64. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Ravera S, Reyna-Neyra A, Ferrandino G, Amzel LM, Carrasco N. The sodium/iodide symporter (NIS): molecular physiology and preclinical and clinical applications. *Annu Rev Physiol.* 2017;79:261-89. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Bloise FF, Cordeiro A, Ortega-Carvalho TM. Role of thyroid hormone in skeletal muscle physiology. *J Endocrinol.* 2018;236(1):R57-68. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Biondi B. Subclinical hypothyroidism in patients with obesity and metabolic syndrome: a narrative review. *Nutrients.* 2023;16(1):87. [View at Publisher] [DOI] [PMID] [Google Scholar]

5. Lewandowski MJ, Krenn S, Kurnikowski A, Bretschneider P, Sattler M, Schwaiger E, et al. Chronic kidney disease is more prevalent among women but more men than women are under nephrological care: analysis from six outpatient clinics in Austria 2019. *Wien Klin Wochenschr*. 2023;135(3-4):89-96. [View at Publisher] [DOI] [PMID] [Google Scholar]
6. Zúñiga D, Balasubramanian S, Mehmood KT, Al-Baldawi S, Salazar GZ. Hypothyroidism and Cardiovascular Disease: A Review. *Cureus*. 2024;16(1):e52512. [View at Publisher] [DOI] [PMID] [Google Scholar]
7. Ahmad SS, Chun HJ, Ahmad K, Shaikh S, Lim JH, Ali S, et al. The roles of growth factors and hormones in the regulation of muscle satellite cells for cultured meat production. *J Anim Sci Technol*. 2023;65(1):16-31. [View at Publisher] [DOI] [PMID] [Google Scholar]
8. Sabatino L. Nrf2-Mediated antioxidant defense and thyroid hormone signaling: a Focus on Cardioprotective effects. *Antioxidants*. 2023;12(6):1177. [View at Publisher] [DOI] [PMID] [Google Scholar]
9. Fasciolo G, Napolitano G, Aprile M, Cataldi S, Costa V, Muscari Tomajoli MT, et al. Muscle oxidative stress plays a role in hyperthyroidism-linked insulin resistance. *Antioxidants*. 2023;12(3):592. [View at Publisher] [DOI] [PMID] [Google Scholar]
10. Machado M, Bachini F, Itaborahy A. Thyroid Hormones and Skeletal Muscle Beyond Thermogenesis. *J Sci Sport Exerc*. 2023;6:315-23. [View at Publisher] [DOI] [Google Scholar]
11. Agostini D, Gervasi M, Ferrini F, Bartolacci A, Stranieri A, Piccoli G, et al. An integrated approach to skeletal muscle health in aging. *Nutrients*. 2023;15(8):1802. [View at Publisher] [DOI] [PMID] [Google Scholar]
12. Gu X, Wang L, Liu S, Shan T. Adipose tissue adipokines and lipokines: Functions and regulatory mechanism in skeletal muscle development and homeostasis. *Metabolism*. 2023;139:155379. [View at Publisher] [DOI] [PMID] [Google Scholar]
13. Drabińska N, Romaszko J, White P. The effect of isocaloric, energy-restrictive, KETOgenic diet on metabolism, inflammation, nutrition deficiencies and oxidative stress in women with overweight and obesity (KETO-MINOX): Study protocol. *Plos One*. 2023;18(5):e0285283. [View at Publisher] [DOI] [PMID] [Google Scholar]
14. Chen J, Wei L, Zhu X, Xu W, Zou Y, Qi X, et al. TT3, a more practical indicator for evaluating the relationship between sarcopenia and thyroid hormone in the euthyroid elderly compared with FT3. *Clin Interv Aging*. 2023;1285-93. [View at Publisher] [DOI] [PMID] [Google Scholar]
15. Miro C, Nappi A, Saggiocchi S, Di Cicco E, Murolo M, Torabinejad S, et al. Thyroid hormone regulates the lipid content of muscle fibers, thus affecting physical exercise performance. *Int J Mol Sci*. 2023;24(15):12074. [View at Publisher] [DOI] [PMID] [Google Scholar]
16. Kaspari RR, Reyna-Neyra A, Jung L, Torres-Manzo AP, Hirabara SM, Carrasco N. The paradoxical lean phenotype of hypothyroid mice is marked by increased adaptive thermogenesis in the skeletal muscle. *Proc Natl Acad Sci U S A*. 2020;117(36):22544-51. [View at Publisher] [DOI] [PMID] [Google Scholar]
17. De Stefano MA, Ambrosio R, Porcelli T, Orlandino G, Salvatore D, Luongo C. Thyroid hormone action in muscle atrophy. *Metabolites*. 2021;11(11):730. [View at Publisher] [DOI] [PMID] [Google Scholar]
18. Nappi A, Moriello C, Morgante M, Fusco F, Crocetto F, Miro C. Effects of thyroid hormones in skeletal muscle protein turnover. *J Basic Clin Physiol Pharmacol*. 2024;35(4-5):253-64. [View at Publisher] [DOI] [PMID] [Google Scholar]
19. Hegedüs L, Bianco AC, Jonklaas J, Pearce SH, Weetman AP, Perros P. Primary hypothyroidism and quality of life. *Nat Rev Endocrinol*. 2022;18(4):230-42. [View at Publisher] [DOI] [PMID] [Google Scholar]
20. Mirzaei A, Mirzaei N, Ghavamizadeh M. Antioxidant activity and cytotoxicity of *Dorema aucheri* by *Artemia urmiana*: a brine shrimp lethality test. *Life Sci J*. 2013;10:8-12. [View at Publisher] [Google Scholar]
21. Di Dalmazi G, Giuliani C. Plant constituents and thyroid: A revision of the main phytochemicals that interfere with thyroid function. *Food Chem Toxicol*. 2021;152:112158. [View at Publisher] [DOI] [PMID] [Google Scholar]
22. Hajibabae F, Abedpoor N, Safavi K, Taghian F. Natural remedies medicine derived from flaxseed (secoisolariciresinol diglucoside, lignans, and  $\alpha$ -linolenic acid) improve network targeting efficiency of diabetic heart conditions based on computational chemistry techniques and pharmacophore modeling. *J Food Biochem*. 2022;46(12):e14480. [View at Publisher] [DOI] [PMID] [Google Scholar]
23. Hajibabae F, Abedpoor N, Taghian F, Safavi K. A cocktail of polyherbal bioactive compounds and regular mobility training as senolytic approaches in age-dependent alzheimer's: the in silico analysis, lifestyle intervention in old age. *J Mol Neurosci*. 2023;73(2-3):171-84. [View at Publisher] [DOI] [PMID] [Google Scholar]
24. Mostafavi SH, Fazilati M, Mostafavi S-A, Vahhabi MR, Mostafavi F, Omidvarinia S, et al. Hepatotoxicity of *Dorema aucheri* (Bilhar) in albino mice. *Arch Iran Med*. 2013;16(9):530-2. [View at Publisher] [Google Scholar]
25. Ismail RF, Hamed M, Sayed AE-DH. Lycopene supplementation: effects on oxidative stress, sex hormones, gonads and thyroid tissue in tilapia *Oreochromis niloticus* during Harness® exposure. *Front Physiol*. 2023;14:1237159. [View at Publisher] [DOI] [PMID] [Google Scholar]
26. Arida RM, de Almeida A-CG, Cavalheiro EA, Scorza FA. Experimental and clinical findings from physical exercise as complementary therapy for epilepsy. *Epilepsy Behav*. 2013;26(3):273-8. [View at Publisher] [DOI] [PMID] [Google Scholar]
27. Izquierdo M, Singh MF. Promoting resilience in the face of ageing and disease: The central role of exercise and physical activity. *Ageing Res Rev*. 2023;88:101940. [View at Publisher] [DOI] [PMID] [Google Scholar]
28. Bennett S, Sato S. Enhancing the metabolic benefits of exercise: Is timing the key? *Front Endocrinol (Lausanne)*. 2023;14:987208. [View at Publisher] [DOI] [PMID] [Google Scholar]
29. Ayhan S, Tizar E, Erdoğan R. Investigation of the Effect of High-Intensity Training on Mineral and Thyroid Hormone Metabolism of Athletes. *Journal of Education and Recreation Patterns*. 2023;4(2):275-82. [View at Publisher] [DOI] [Google Scholar]
30. Boström P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, et al. A PGC1- $\alpha$ -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature*. 2012;481(7382):463-8. [View at Publisher] [DOI] [PMID] [Google Scholar]
31. Huh JY, Mougios V, Kabasakalis A, Fatouros I, Siopi A, Douroudos II, et al. Exercise-induced irisin secretion is independent of age or fitness level and increased irisin may directly modulate muscle metabolism through AMPK activation. *J Clin Endocrinol Metab*. 2014;99(11):E2154-61. [View at Publisher] [DOI] [PMID] [Google Scholar]
32. Samy DM, Ismail CA, Nassra RA. Circulating irisin concentrations in rat models of thyroid dysfunction-effect of exercise. *Metabolism*. 2015;64(7):804-13. [View at Publisher] [DOI] [PMID] [Google Scholar]
33. Singh S, Panda V, Sudhamani S, Dande P. Protective effect of a polyherbal bioactive fraction in propylthiouracil-induced thyroid toxicity in rats by modulation of the hypothalamic-pituitary-thyroid and hypothalamic-pituitary-adrenal axes. *Toxicol Rep*. 2020;7:730-42. [View at Publisher] [DOI] [PMID] [Google Scholar]
34. Hossiniyan SA, Latifpour M, Mohammadi B, Jadidi-Niaragh F, Ganji A, Ghalamfarsa G, et al. Chemical composition and cytotoxic activity of the essential oil from the aerial parts of *Dorema aucheri*. *J Herbm Pharm*. 2021;10(3):344-50. [View at Publisher] [DOI] [Google Scholar]
35. Abedpoor N, Taghian F, Ghaedi K, Niktab I, Safaeinejad Z, Rabiee F, et al. PPAR $\gamma$ /Pgc-1 $\alpha$ -Fndc5 pathway up-regulation in gastrocnemius and heart muscle of exercised, branched chain amino acid diet fed mice. *Nutr Metab (Lond)*. 2018;15:59. [View at Publisher] [DOI] [PMID] [Google Scholar]
36. Hu X, Sun M, Chen Q, Zhao Y, Liang N, Wang S, et al. Skeletal muscle-secreted DLPC orchestrates systemic energy homeostasis by enhancing adipose browning. *Nat Commun*. 2023;14(1):7916. [View at Publisher] [DOI] [PMID] [Google Scholar]
37. Seke Etet PF, Vecchio L, Kamdje AHN, Mimche PN, Njamnshi AK, Adem A. Physiological and environmental factors affecting cancer risk and prognosis in obesity. *Semin Cancer Biol*. 2023;94:50-61. [View at Publisher] [DOI] [PMID] [Google Scholar]
38. Furrer R, Hawley JA, Handschin C. The molecular athlete: exercise physiology from mechanisms to medals. *Physiol Rev*. 2023;103(3):1693-787. [View at Publisher] [DOI] [PMID] [Google Scholar]
39. Salvatore D, Simonides WS, Dentice M, Zavacki AM, Larsen PR. Thyroid hormones and skeletal muscle-new insights and potential implications. *Nat Rev Endocrinol*. 2014;10(4):206-14. [View at Publisher] [DOI] [PMID] [Google Scholar]
40. Benzie IFF, Wachtel-Galor S, eds. *Herbal medicine: biomolecular and clinical aspects*. 2nd ed. Boca Raton (FL): CRC Press/Taylor & Francis; 2011. [View at Publisher] [PMID] [Google Scholar]
41. Rasouli Vani J, Taghi Mohammadi M, Sarami Foroshani M, Rezazade E. Evaluation of the neuroprotective and antioxidant effects of *Dorema aucheri* extract on cerebral ischaemia-reperfusion injury in rats. *Pharm Biol*. 2019;57(1):255-62. [View at Publisher] [DOI] [PMID] [Google Scholar]
42. Mianabadi M, Hoshani M, Salmanian S. Antimicrobial and Anti-oxidative Effects of Methanolic Extract of *Dorema aucheri* Boiss. *J Agric Sci Technol*. 2015;17(3):623-34. [View at Publisher] [Google Scholar]
43. Coppola A, Liu Z-W, Andrews ZB, Paradis E, Roy M-C, Friedman JM, et al. A central thermogenic-like mechanism in feeding regulation: an interplay between arcuate nucleus T3 and UCP2. *Cell Metab*. 2007;5(1):21-33. [View at Publisher] [DOI] [PMID] [Google Scholar]
44. Bray G. Reciprocal relation of food intake and sympathetic activity: experimental observations and clinical implications. *Int J Obes Relat Metab Disord*. 2000;24(Suppl 2):S8-17. [View at Publisher] [DOI] [PMID] [Google Scholar]
45. Racotta IS, Cabanac M, Lafrance L. Dissociation of anorectic and affective responses to epinephrine and glucose in rats. *Physiol Behav*. 1995;58(1):125-30. [View at Publisher] [DOI] [PMID] [Google Scholar]

46. Sanchez NM, Varela L, Martins L, Morentin PM, Gallego R, Vazquez M, et al, eds. Hypothalamic mTOR pathway mediates thyroid hormone-induced hyperphagia in hyperthyroidism. *Endocrine Abstracts*. 2012;29:P1564. [[View at Publisher](#)] [[Google Scholar](#)]
47. Kunz D, Oster H, Rawashdeh O, Neumann WJ, Münte T, Berg D. Sleep and circadian rhythms in  $\alpha$ -synucleinopathies-perspectives for disease modification. *Acta Physiol (Oxf)*. 2023;238(1):e13966. [[View at Publisher](#)] [[DOI](#)] [[PMID](#)] [[Google Scholar](#)]
48. Sohrabi O, Hatamzadeh A, Ghasemnezhad A, Samizadeh H, Erfani-Moghadam V. Exploring the effects of Medicinal Plant extracts on Tomato (*Solanum lycopersicum* L.) morphology, Biochemistry, and Plant Growth regulators under greenhouse conditions. *Int J Horti Sci Technol*. 2024;11(3):285-98. [[View at Publisher](#)] [[DOI](#)] [[Google Scholar](#)]
49. Fortunato RS, Ignacio DL, Padron AS, Peçanha R, Marassi MP, Rosenthal D, et al. The effect of acute exercise session on thyroid hormone economy in rats. *J Endocrinol*. 2008;198(2):347-54. [[View at Publisher](#)] [[DOI](#)] [[PMID](#)] [[Google Scholar](#)]
50. Mullur R, Liu Y-Y, Brent GA. Thyroid hormone regulation of metabolism. *Physiol Rev*. 2014;94(2):355-82. . [[View at Publisher](#)] [[DOI](#)] [[PMID](#)] [[Google Scholar](#)]
51. Csaba G. Hormones in the immune system and their possible role. A critical review. *Acta Microbiol Immunol Hung*. 2014;61(3):241-60. [[View at Publisher](#)] [[DOI](#)] [[PMID](#)] [[Google Scholar](#)]

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